

Lecture #26

Lecture 26

4/16/04

No class Monday(patriots day) OR Wednesday(4/21)

New X-ray structure of the translocon:

Nature (2004) 427, 36-44

Targeting of proteins to membranes and for secretion

p.2 Handout 4a

Cartoon overview of SRP pathway

Equipment required for inserting proteins into membranes

Both SRP and SRP-R contain GTPase domains

Somehow (we don't completely understand the whole process yet) SRP interacts with signal sequence (hydrophobic) at N-terminus of growing polypeptide as it exits the ribosome, a pause in polypeptide formation occurs because the SRP can interact with the A site of the ribosome. The pause allows time for the SRP with its cargo to interact with SRP-R in the membrane, and transports protein to translocon (ER membrane)

How are multiple transmembrane domains inserted? We don't know yet.

Translocon x-ray structure does not agree with other methods (FRET, cryoEM)

We need multiple methods to really understand function

How to get proteins into the mitochondria?

p.4 handout 4a

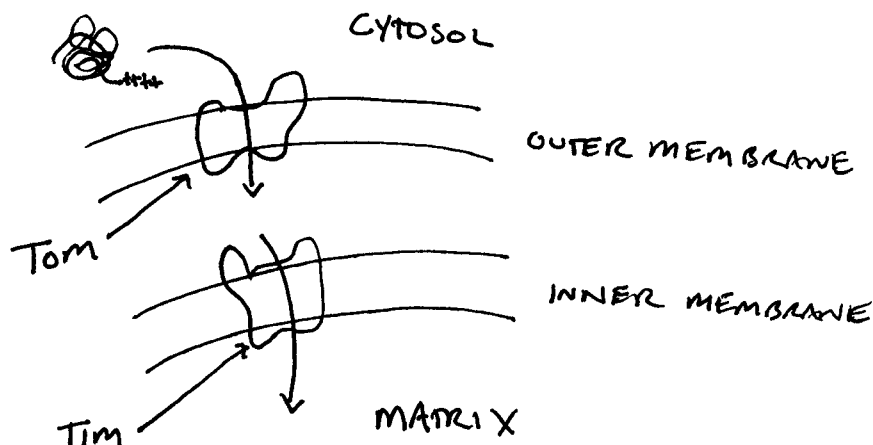
Must cross inner and outer membrane to get into mitochondrial matrix

+++++, a positively charged signal sequence at the N-terminus (first end to exit ribosome), targets the protein for transport into the mitochondria

TOM (transporter outer membrane) complex of proteins- equipment to get protein across outer membrane

TIM (transporter inner membrane) complex of proteins- equipment to get protein across inner membrane

Cartoon Diagram:



Delta Psi, a change in membrane potential is required for transport across the inner mitochondrial membrane.

p.4 handout 4a

To go from cytosol into matrix you have to unfold and then refold the protein
Why do this?

3 possibilities for how folding and refolding could be accomplished

- a. spontaneous global unfolding
(not as far-fetched as it sounds, the energy difference between folded/unfolded states is very small)
- b. catalyzed unfolding – membrane potential
- c. catalyzed unfolding – chaperone protein (uses ATP)

Misfolding:

We know that misfolding occurs

Can we refold misfolded proteins? Yes

If we cannot correct the misfolding, can we remove the misfolded protein? Yes, via the proteasome

If we can't correct or destroy the misfolded protein, what are the consequences?

-disease, (ex. Mad cow disease, alzheimer's, huntington's disease, parkinson's)

See p. 5 handout 4a

Misfolded proteins can form amyloid plaques and other aggregates

Protein Folding *in vitro*

Energy difference between folded/unfolded state is very small (~5-15 kcal/mol)

A typical H-bond is ~5 kcal/mol

People would love to be able to predict protein folded structure from primary sequence, but very difficult problem

There are huge numbers of interactions in both the folded and unfolded states, and the difference between the overall energies is very small- makes prediction of protein folding very difficult

The Levinthal Paradox

Proteins cannot undergo unbiased conformational space searches to fold

If a protein were to sample all the different conformational space available it would take billions of years for proteins to fold!

Need a biased search.

Anfinsen's Hypothesis: The primary sequence is sufficient for protein folding

Did experiments on RNase (ribonuclease) – cleaves RNA

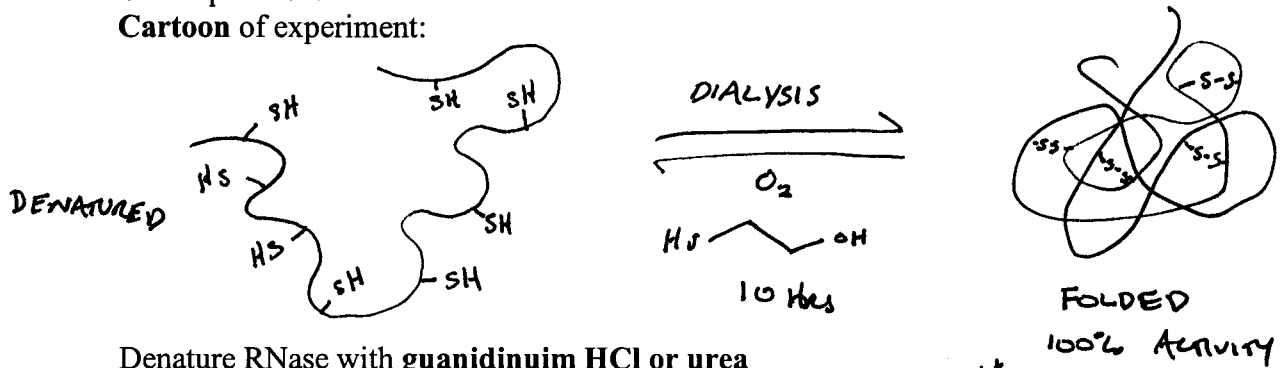
124 amino acid protein

RNase was chosen because it was available in gram quantities, also the protein folding can be easily measured by catalytic activity (very sensitive)

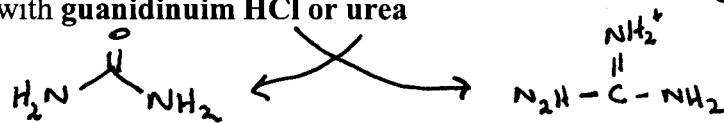
RNase has 8 cysteines-and the active form of the protein contains four disulfides. How do you form the correct disulfide bonds?

One experiment:

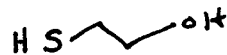
Cartoon of experiment:



Denature RNase with guanidinium HCl or urea



Then dialyze away the denaturant in the presence of **beta-mercapethanol (beta-Me)**
 After 10 h the unfolded protein is refolded as evidenced by the recovery of 100% of its catalytic activity



Dialysis without (beta-Me) and in the presence of oxygen gives only 1% activity

Why only 1% activity w/out beta-Me? The chance of randomly forming the correct disulfide bonds on the first go is very small

1 in 7 chance of forming first bond correctly, for the next bond you have a 1 in 5 chance, etc

$1/7 \times 1/5 \times 1/3 = 1/105$ chance forming all four disulfide bonds with no corrections

Therefore, only 1% of the protein is folded correctly and is active

Beta-Me

Allows disulfide bonds to form, break and reform

Protein can sample different disulfide bond combinations, eventually get 100% folded and active protein

Cartoon of Beta-Me allowing disulfides to break and reform

